

****PLEASE CREATE A STERILE WORK STATION AND BE MASKED & GLOVED BEFORE PROCEEDING****

Wipe sealing port of anticoagulant and heparin with sterile alcohol prior to accessing with a sterile needle/syringe

For questions please contact:
844-897-4910

Step 1:



Draw 20mL of Heparin into 60mL Syringe

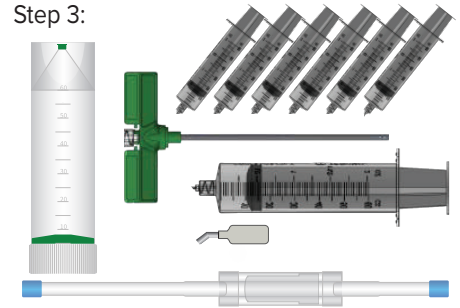
Step 2:

Add the female-to-female connector, use the 60cc syringe to Heparinize the following components:

- (6) 10cc Syringes
- *Leaving 1cc of Heparin in each one*
- (1) 60cc Syringe
- 150um Filter
- Bone Marrow Needle
- 45 Degree Dispensing Tip
- XCELL Concentrating Device

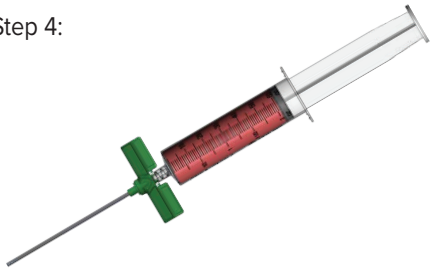


Step 3:



Heparinize 60cc syringe before disposing of remaining Heparin. Prepare a clean working surface and layout all Heparinized contents

Step 4:



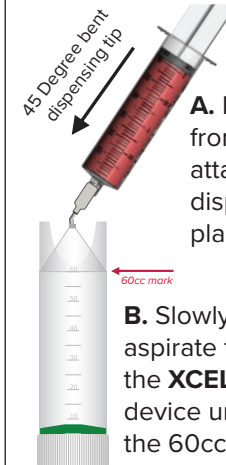
Attach one 10cc syringe to the inserted needle and slowly aspirate marrow to the 10cc mark. Cap syringe and set aside. Repeat with 5 additional 10cc syringes until the desired 60cc amount has been collected.

Step 5:



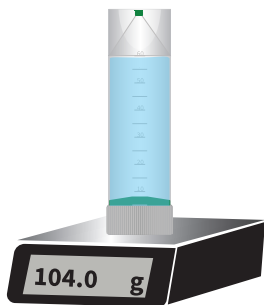
- A. Attach one of the Heparinized 60cc syringes to the 150um filter.
- B. Remove cap from one of the 10cc syringes of bone marrow aspirate and connect to the opposite end of the 150um filter. Pull back slowly on the 60cc syringe pulling the bone marrow through the filter into itself.
- C. Repeat with the remaining (5) 10cc syringes.

Step 6:



- A. Detach the filter from 60cc syringe and attach the 45-degree dispensing tip in its place.
- B. Slowly transfer the aspirate from the syringe into the **XCELL** concentrating device until you've reached the 60cc mark.

Step 7:



**Secure the green silicone stopper and the clear safety cap to the concentrating device. Match counterbalance to +/- 1.0g of concentrating device.

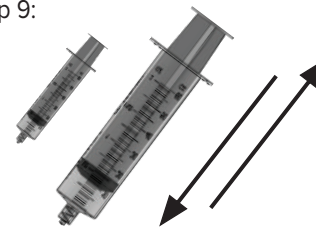
Step 8:

Place **XCELL** counterbalance and concentrating device on opposite ends inside centrifuge and spin:

Drucker:
3900 RPM/2850 RCF
12 minutes

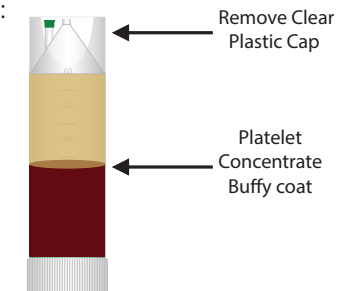
Eppendorf:
4200 RPM/2800 RCF
12 minutes

Step 9:

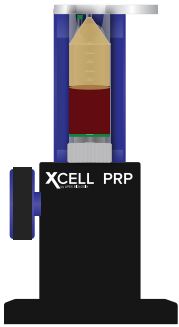
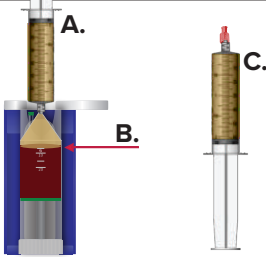
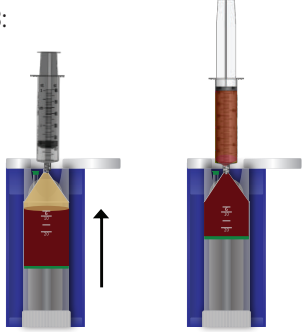
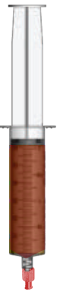


Prime a 60mL and 10mL syringe to ensure that the barrel moves freely. This is done by simply pulling back and forth on the plunger two to three times. Leave 5mL of air in the 60mL syringe to prevent splatter

Step 10:



After spin, carefully remove **XCELL** concentrating device from the centrifuge. Remove the caps from Step 4

<p>Step 11:</p>  <p>Place the Concentrating Device into the Bench Top Processing Station and slowly turn the knob until the bone marrow aspirate has reached the bottom of the luer slip fitting.</p>	<p>Step 12:</p>  <p>A. Place primed 60mL Syringe vertically on XCELL concentrating device B. Using the Bench Top Processing Station push PPP into 60mL syringe until the buffy coat reaches 6mL (outlined on concentrating device.) (See red arrow) C. Remove and cap 60mL syringe</p>	<p>Step 13:</p>  <p>Aspirate BMC</p> <p>***Keeping the assembly vertical, add the primed 10ml syringe and push the remaining BMC until the syringe captures the buffy coat</p>	<p>Step 14:</p>  <p>Cap the 10ml syringe and gently remix the suspension. The XCELL BMC process is complete</p>
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***This process provides 6-6.5ml concentrate. For higher TNC counts, continue pushing RBC into 10cc syringe to the 8-9ml mark. If lower volume is desired, push the buffy coat above the 6ml marking on the Concentrating Device in step 12, add the 10cc syringe, and push in the desired volume.